

Preliminary communication

Synthetic studies on rhynchosporoside: Stereoselective synthesis of 1-*O*- and 2-*O*- α -cellobiosyl-3-deoxy-2(*R*)- and -2(*S*)-glycerol*

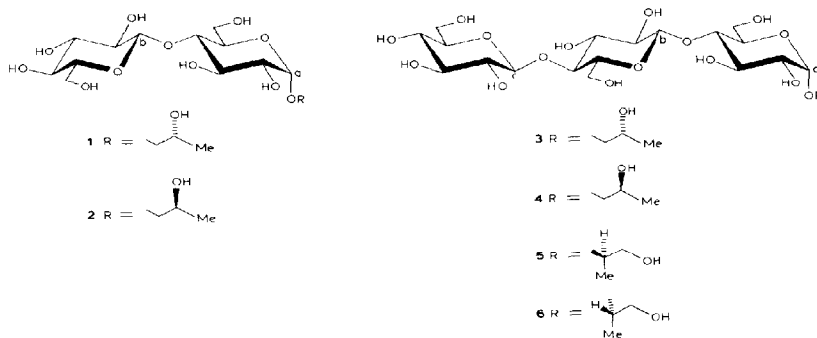
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In 1980, it was proposed² that the structure of rhynchosporoside, a phytotoxic compound, is 1-*O*- α -cellobiosyl- or 1-*O*- α -cellobiosyl-3-deoxyglycerol and in 1982, we reported¹ that 1-*O*- α -cellobiosyl-3-deoxy-2(*R*)-glycerol (**1**) is more toxic towards *Hordeum sativum* Jessen than the 2(*S*) isomer **2**.

We now describe a stereoselective synthesis of the α -cellobiosyl derivatives **3–6**. A preliminary biological test[†] showed that **3** is more toxic, and **5** and **6** are less toxic, than **1**, whereas **4** is not toxic at all as in the case of **2**. These results indicated that both the 2(*R*) configuration of the diol residue and the α -D configuration of the cellobiosyl group on the primary hydroxyl group of the diol are required for production of the phytotoxicity.



*Synthetic Studies on Glycosidic Phytotoxins, Part II. Fort Part I, see ref. 1.

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[†]Phytotoxicity of compounds **3–6** towards *Hordeum sativum* Jessen was examined by us. Prof.

Strobel performed biological testing of **1–4** towards *Hordeum vulgare* L., and observed the same trend for the toxicity of the synthetic samples as did we. We are grateful to Prof. Strobel for his kind co-operation.

In planning the synthesis of **3**, **4**, **5**, and **6**, the target structures were retrosynthesized into the glycosyl acceptors **8**, **9**, **10**, or **11**, the D-glucosyl donors **13** or **14**, and the cellobiosyl donor **19**. The chiral glycosyl acceptors **8–11** were prepared by starting from 3-deoxy-2(*S*)-glycerol (**7**), readily available from L-(+)-lactic acid by LiAlH_4 reduction. Compound **7** was converted into **8** in 5 steps (44%); namely (i) TrCl , (ii) $\text{Ph}_3\text{P} \cdot \text{PhCO}_2\text{H} \cdot \text{DEAD}^5$ in THF, (iii) $\text{NaOMe} \cdot \text{MeOH}$, (iv) $\text{BnBr} \cdot \text{NaH}$ in DMF, and (v) camphorsulfonic acid (CSA) in MeOH; the 3,5-dinitrobenzoate (DNB) of **8** had $[\alpha]_D -18.3^\circ$. Compound **9** was prepared in 3 steps (65%) from **7**: (i) TrCl , (ii) $\text{BnBr} \cdot \text{NaH}$ in DMF, and (iii) CSA in MeOH; 3,5-DNB of **9**, $[\alpha]_D +17.8^\circ$. Compound **11** was prepared from **7** in 70% yield: (i) $(\text{Bu}_3\text{Sn})_2\text{O}^4$, and (ii) $\text{BnBr} \cdot \text{Bu}_4\text{NB}^6$, 3,5-DNB of **11**, $[\alpha]_D +29.0^\circ$. Compound **10** was prepared from **11** in 87% yield: (i) $\text{Ph}_3\text{P} \cdot \text{PhCO}_2\text{H} \cdot \text{DEAD}$ and (ii) $\text{NaOMe} \cdot \text{MeOH}$; 3,5-DNB of **10**, $[\alpha]_D -29.1^\circ$.

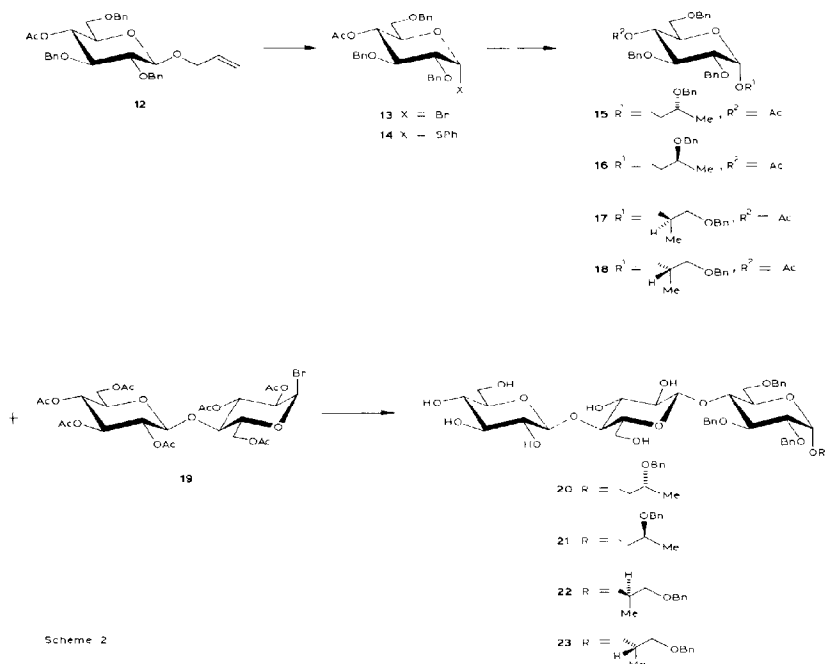


By using these chiral propanediol derivatives as the glycosyl acceptors, the target molecules **3** and **4** were synthesized in the following way. Allyl β -D-glucopyranoside **12**¹ was treated with (i) $\text{PdCl}_2 \cdot \text{AcONa} \cdot \text{aq. AcOH}^6$ and (ii) $\text{Ph}_3\text{P} \cdot \text{CBr}_4$ ⁷, to give **13**, which was respectively treated with the glycosyl acceptors **8** and **9** in the presence of $\text{Et}_4\text{NB} \cdot \text{powdered molecular sieves}^8$ **4A** in $\text{Cl}(\text{CH}_2)_2\text{Cl}$ for 3 days at 70° , to give **15**, $[\alpha]_D +29.2^\circ$, and **16**, $[\alpha]_D +41.0^\circ$, in 47 and 32% yield, respectively. Compound **15** was then converted into **20**, $[\alpha]_D +39.1^\circ$ (R_F 0.45 in 9:1 $\text{CHCl}_3 \cdot \text{MeOH}$), in 3 steps (55%): (i) $\text{NaOMe} \cdot \text{MeOH}$, (ii) **19**– $\text{AgOSO}_2\text{CF}_3$ –powdered molecular sieves **4A** in $\text{Cl}(\text{CH}_2)_2\text{Cl}$, and (iii) $\text{NaOMe} \cdot \text{MeOH}$. The same reaction-sequence with **16** afforded a 63% yield of **21**, $[\alpha]_D +25.0^\circ$; R_F 0.45 in 9:1 $\text{CHCl}_3 \cdot \text{MeOH}$. Catalytic hydrogenolysis of **20** and **21** in the presence of 10% $\text{Pd} \cdot \text{C}$ in MeOH at 50° respectively afforded **3**, $[\alpha]_D +58.0^\circ$ (H_2O); R_F 0.30 in 20:20:1 $\text{CHCl}_3 \cdot \text{MeOH} \cdot \text{AcOH}$; δ_C (D_2O): 103.25 and 103.08 (C-1b and C-1c, $^1J_{\text{CH}}$ 162.1 Hz), 99.10 (C-1a, $^1J_{\text{CH}}$ 170.9 Hz), 79.33 and 79.17 (C-4a and C-4b), and 18.81 (CH_3), and **4**, $[\alpha]_D +65.0^\circ$ (H_2O); R_F 0.30 in 20:20:1 $\text{CHCl}_3 \cdot \text{MeOH} \cdot \text{AcOH}$; δ_C (D_2O): 103.35 and 103.08 (C-1b and C-1c, $^1J_{\text{CH}}$ 163.1 Hz), 98.55 (C-1a, $^1J_{\text{CH}}$ 169.9 Hz), 79.33 and 79.17 (C-4a and C-4b), and 18.82 (CH_3).

Next, the target molecules **5** and **6**, the regio-isomers of **3** and **4**, were synthesized as follows. Allyl β -D-glucopyranoside (**12**) was converted into a mixture of **14**,

[†] Values of $[\alpha]_D$ were measured for CHCl_3 solutions at 25° , unless noted otherwise. Compounds having $[\alpha]_D$ recorded gave satisfactory data for elemental analyses.

[§] The values of δ_C are expressed in p.p.m. downward from tetramethylsilane, referenced indirectly with an internal standard of 1,4-dioxane (δ_C 67.10).



Scheme 2

δ_C (CDCl₃): 87.1 (C-1), δ_H (CDCl₃): 1.88 (Ac), and its β anomer, δ_C (CDCl₃): 87.7 (C-1), δ_H (CDCl₃): 1.86 (Ac), in the ratio of 3:4, in 3 steps (72% yield): (i) PdCl₂-AcONa-aq.AcOH, (ii) Ac₂O-pyridine, and (iii) Bu₃SnSPh-Me₃SiOSO₂CF₃ in CH₂Cl₂⁹. The glycosylation of **10** with a mixture of **14** and its β anomer in the presence of NBS and powdered molecular sieves **4A**¹⁰, and deacetylation of the product, afforded a 43% yield of **17**, $[\alpha]_D +52.4^\circ$; R_F 0.36 in 5:1 toluene-EtOAc, and a 44% yield of the β anomer of **17**, $[\alpha]_D -15.3^\circ$; R_F 0.30 in 5:1 toluene-EtOAc. In the same way, **11** was converted into **18**, $[\alpha]_D +39.1^\circ$; R_F 0.36 in 5:1 toluene-EtOAc, and the β anomer of **18**, $[\alpha]_D -11.4^\circ$; R_F 0.30 in 5:1 toluene-EtOAc, in 25 and 31% yield respectively. Further transformation of **17** and **18** into **5** and **6**, respectively, was achieved *via* **22**, $[\alpha]_D +50.0^\circ$ (R_F 0.45 in 9:1 CHCl₃-MeOH) and **23**, $[\alpha]_D +37.5^\circ$ (R_F 0.45), in 31 and 30% yield, respectively, as in the case of **15** and **16**; **5**, $[\alpha]_D +44.3^\circ$ (H₂O), R_F 0.30, **6**, $[\alpha]_D +62.0^\circ$ (H₂O), R_F 0.30 in 20:20:1 CHCl₃-MeOH-AcOH.

Comparison of the ¹H-n.m.r. data for the samples of synthetic **3**, **4**, **5**, and **6** (see Table I) with those reported² for rhynchosporoside did not definitely identify the structure of the natural product, owing to the presence, in the spectrum of the natural

TABLE I

400-MHz, ^1H -N.M.R. DATA FOR SYNTHETIC COMPOUNDS ^a

Compound	H-1c	H-1b	H-1a	CH ₃
1		4.521 (7.81)	4.936 (3.91)	1.181 (6.35)
2		4.523 (7.81)	4.939 (3.90)	1.202 (6.35)
3	4.517 (7.81)	4.545 (8.30)	4.935 (3.42)	1.180 (6.35)
4	4.517 (7.81)	4.546 (8.30)	4.938 (3.42)	1.202 (6.84)
5	4.517 (7.81)	4.547 (7.81)	5.097 (3.91)	1.234 (6.35)
6	4.516 (7.81)	4.546 (8.06)	5.063 (3.91)	1.166 (6.35)

^a The values of δH are expressed in p.p.m. downward from the internal standard, sodium 2,2,3,3-tetradeterio-4,4-dimethyl-4 silapentanoate, in D_2O . The values in parentheses are $^3\text{J}_{\text{HH}}$ values expressed in Hz.

sample, of a broad DOH signal at δ 4.7–5.0 which was overlapped by the signals of H-1a of the proposed structures 3 and 4.

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